

Regio- and Diastereoselective Synthesis of Highly Substituted, Oxygenated Piperidines from Tetrahydropyridines

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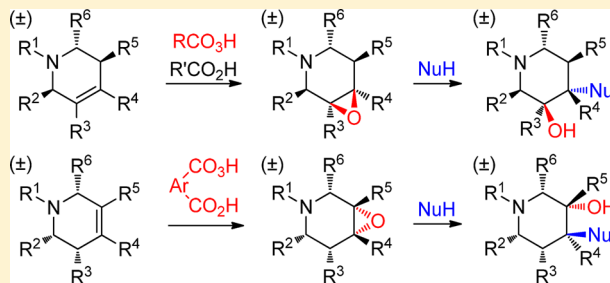
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Supporting Information

ABSTRACT: Diastereoselective epoxidation and regioselective ring-opening methods were developed for the synthesis of densely substituted, oxygenated piperidines from two classes of tetrahydropyridines with distinct stereochemical displays of functionalities. A new and practical in situ prepared epoxidation reagent was developed for the diastereoselective epoxidation of one class of sterically hindered tetrahydropyridines. The novel bifunctional epoxidation reagent, 2-carboperoxy-3,4,5,6-tetrafluorobenzoic acid, was designed to incorporate highly reactive percarboxy acid and pendant carboxylic acid groups, which through hydrogen bonding to the amino group successfully overrode steric effects and directed epoxidation to occur at the more hindered face of the tetrahydropyridine. Nucleophilic ring-opening of the epoxides with water, alcohols, and HF proceeded with high regioselectivity, affording piperidinol products with adjacent tetrasubstituted carbons.



INTRODUCTION

Piperidine rings are ubiquitous in natural products and pharmaceuticals that play pivotal roles in the treatment of disease.^{1–3} For this reason, various methods have been developed to prepare piperidines, although few methods enable the efficient preparation of densely substituted derivatives with high levels of regio- and stereocontrol.^{4,5}

We previously disclosed high-yielding and diastereoselective one-pot syntheses of densely substituted tetrahydropyridines **2** and **3** from simple imine and alkyne precursors (Scheme 1a). These tetrahydropyridines, which have different alkene regiochemistries and stereochemical displays, are obtained by a Rh(I)-catalyzed C–H alkenylation/electrocyclization cascade to give a common 1,2-dihydropyridine intermediate **1** followed by divergent kinetic or thermodynamic protonation and reduction sequences.^{6,7}

While this convergent tetrahydropyridine synthesis approach provides rapid access to tetrahydropyridines, the preparation of piperidines requires further elaboration of the alkene functionality. Stereoselective epoxidation of the alkene group in **2** and **3** would arguably be the most powerful and versatile transformation because subsequent nucleophilic ring opening would enable the introduction of diverse functionalities within a drug relevant piperidinol framework.

Epoxidation of alkene substrates that contain basic amino functionalities faces the intrinsic chemoselectivity challenge of undesired electrophilic oxidation at nitrogen. To avoid N-oxide formation, acids have been added to protonate and thereby protect the amino group.^{8,9} The resulting ammonium group is

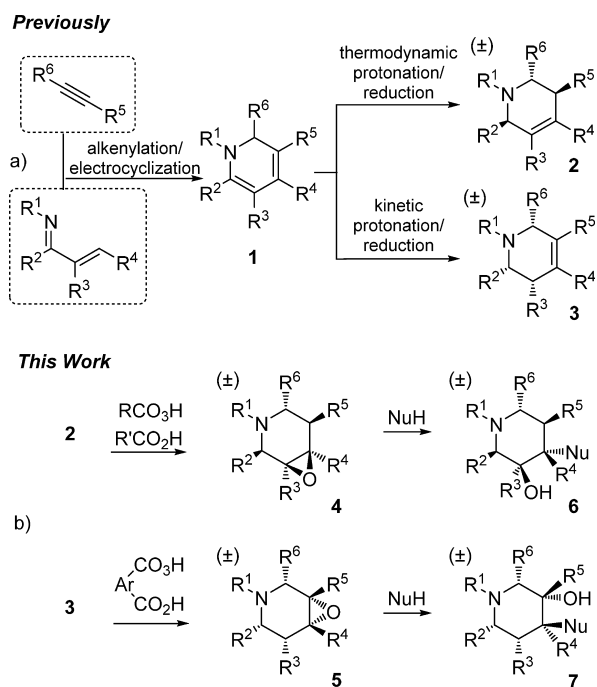
also capable of hydrogen bonding to the peracid to achieve high levels of stereoselectivity through directed epoxidation.

We report here our exploration of diastereoselective epoxidation of **2** and **3**. We find that tetrahydropyridines **2** obtained from a thermodynamic protonation/reduction sequence undergo highly diastereoselective epoxidation of the alkene via protonation and hydrogen bond directed epoxidation (Scheme 1b). However, for tetrahydropyridines **3**, which have a different stereochemical display of substituents about the six-membered ring, this approach resulted in poor epoxidation stereoselectivity. To address this problem, we have developed a new and practical in situ prepared ammonium-directed epoxidation reagent that bears a highly reactive peracid functionality covalently tethered to a carboxylic acid group. Successful highly diastereoselective epoxidation of **3** is achieved by hydrogen bonding of the carboxylic acid group to the amino group, thereby enforcing high face selectivity for epoxidation. This new amino-directed epoxidation reagent should be useful for the oxidation of other alkene substrate classes that contain amino functionalities. Moreover, we demonstrate that nucleophiles react with epoxides **4** and **5** with high regioselectivity to provide densely substituted piperidinol products **6** and **7**, respectively, with each bearing adjacent tetrasubstituted carbons.

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Scheme 1. Previously Disclosed Tetrahydropyridine Synthesis and Comparison with This Work



RESULTS AND DISCUSSION

Stereoselective Epoxidation of Tetrahydropyridines

2. After evaluating different conditions that previously had been reported for amino-directed epoxidation, we elected to use trichloroacetic acid in excess, conditions initially introduced by Davies.⁹ Protonation of tetrahydropyridine **2a** followed by treatment with *m*-chloroperbenzoic acid furnished epoxide **4a** in high yield, with excellent diastereoselectivity and without any *N*-oxide byproduct (Scheme 2). Consistent with previously reported models for ammonium-directed epoxidation, we speculate that the excellent observed diastereoselectivity results from a transition state in which the ammonium proton directs face selectivity by hydrogen bonding to the peroxy acid group oxygens.¹⁰ The depicted structure of protonated **2a** corresponds to that observed in the X-ray crystal structure of the 2,4,6-trinitrobenzenesulfonate salt of **2a**.^{7b}

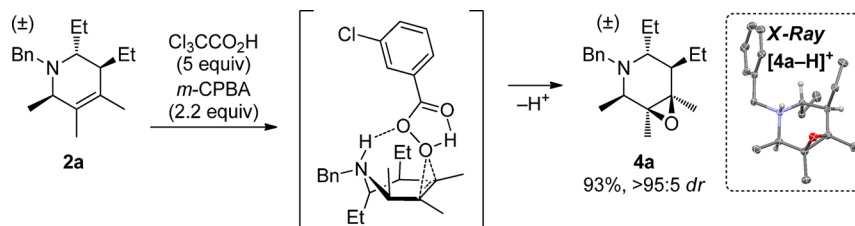
We next explored epoxidation of tetrahydropyridines **2** with a wide range of substitution patterns (vide infra). While the Davies conditions effected complete conversion to the epoxide products for a fair range of tetrahydropyridines with good to excellent diastereoselectivity, we observed unacceptably slow rates of conversion for some derivatives. We therefore sought an alternative epoxidation protocol with an oxidant that is more

reactive than *m*-chloroperbenzoic acid. Trifluoroacetic acid was chosen due to its high reactivity.^{8a,d} Mixing trifluoroacetic anhydride (TFAA) with hydrogen peroxide enabled the in situ preparation of trifluoroacetic acid with concomitant generation of equimolar trifluoroacetic acid to protect the piperidine nitrogen by protonation. Indeed, model substrate **2a** underwent epoxidation cleanly and with high diastereoselectivity (>95:5) at room temperature when treated with premixed TFAA and hydrogen peroxide.

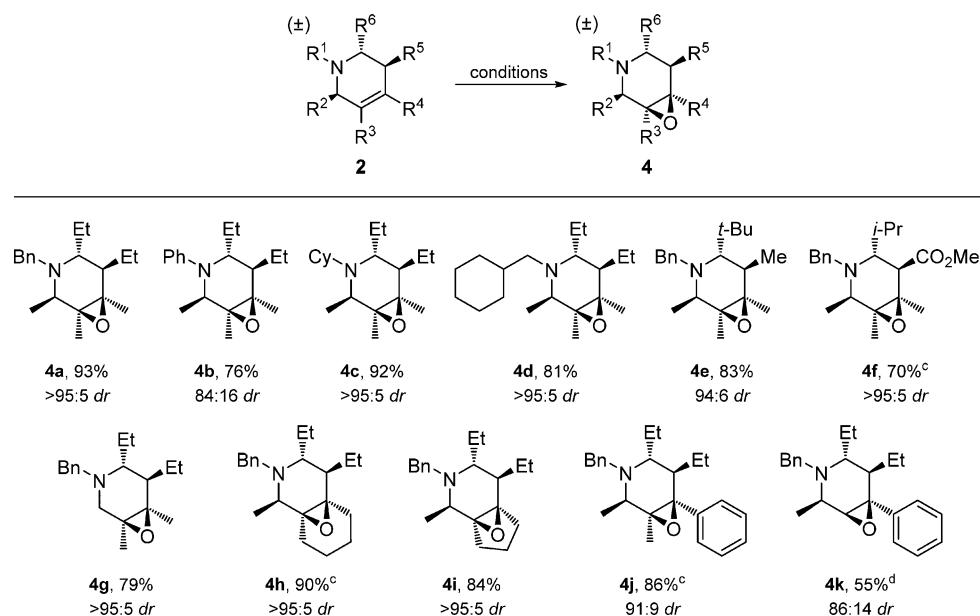
A wide range of tetrahydropyridines **2** derived from the thermodynamic protonation/reduction sequence (Scheme 1) can be successfully converted to epoxides in good to excellent yields and with high diastereoselectivities (Table 1). Different *N*-substitution patterns were well tolerated (**4a–d**). For substrates with deactivating groups (**4f**, **4j**) or sterically hindered substitution patterns (**4h**), TFAA and hydrogen peroxide effected full conversion to the epoxide products in a diastereoselective manner. The relative configuration of the major diastereoisomer was determined unambiguously for epoxides **4a** and **4g** by X-ray crystallographic analysis and assigned by analogy to the rest of the products shown.

Development of a New Class of Peracids for Directed Epoxidation. Tetrahydropyridines **3** display all three substituents R^2 , R^3 , and R^6 on the same side of the heterocycle, which contrasts with tetrahydropyridines **2** with the R^2 and R^5 substituents displayed on the same face and the R^6 substituent on the opposite face of the heterocycle (Scheme 1a). As a result of the different stereochemical display of functionality, the epoxidation conditions that were successful for tetrahydropyridines **2** were ineffective for tetrahydropyridines **3**. For example, epoxidation of **3a** using the mCPBA/ $\text{Cl}_3\text{CCO}_2\text{H}$ conditions proceeded more slowly than for **2a** (see Table 1) with only 77% conversion after 3.5 h and resulted in very poor 51:49 diastereoselectivity. Epoxidation of **3a** with the TFAA and hydrogen peroxide protocol also proceeded with very low diastereoselectivity (Table 2, entry 1). We therefore designed a different hydrogen bonding motif for directly relating face-selectivity of protonation to epoxidation diastereoselectivity. We envisioned that cleaving cyclic or bicyclic anhydrides with hydrogen peroxide would generate a percarboxy acid group tethered to a carboxylic acid group capable of hydrogen bonding to the ammonium proton, thereby controlling the face selectivity of the epoxidation (Scheme 3). In this model, the depicted structure of protonated **3a** corresponds to that observed in the X-ray crystal structure of the 2,4,6-trinitrobenzenesulfonate salt of **3a**.^{7a}

According to this design principle we evaluated a range of cyclic and bicyclic anhydrides (Table 2). Succinic and glutaric anhydrides gave poor conversion (entries 3 and 4), while phthalic anhydride resulted in good conversion but without

Scheme 2. Diastereoselective Epoxidation of Tetrahydropyridine **2a**^a

^aX-ray crystal structure shown with 50% displacement ellipsoids (anion and non-ring hydrogen atoms omitted for clarity).

Table 1. Epoxidation of Tetrahydropyridines 2^{a,b}

^aDiastereoselectivities were determined by ¹H and ¹³C NMR analysis. Isolated yields were determined by mass balance after purification by chromatography. ^bProducts **2a–e**, **2g**, and **2i** were obtained by subjecting the corresponding tetrahydropyridines to Cl₃CCO₂H (5.0 equiv) followed by epoxidation with *m*-CPBA (2.2–3.0 equiv). ^cEpoxidation was accomplished by treating with TFAA (6.0 equiv) and H₂O₂ (30% w/w in H₂O, 5.0 equiv) premixed in THF at 0 °C. ^dThe reported results were obtained by epoxidation with tetrafluorophthalic anhydride and H₂O₂ (vide infra). Epoxidation with TFAA and H₂O₂ resulted in a complex mixture of products.

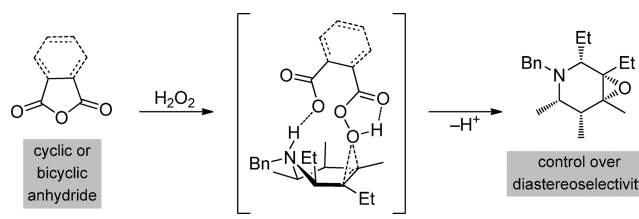
Table 2. Optimization of Epoxidation Conditions for Tetrahydropyridines 3

entry	anhydride	solvent	conversion ^b (%)	dr ^c
1	trifluoroacetic	CH ₂ Cl ₂	100	56:44
2	trifluoroacetic	THF	no reaction	–
3	succinic	CH ₂ Cl ₂	15	58:42
4	glutaric	CH ₂ Cl ₂	27	69:31
5	phthalic	CH ₂ Cl ₂	95	69:31
6	tetrafluorophthalic	CH ₂ Cl ₂	100	42:58
7	tetrafluorophthalic	EtOAc	100	76:24
8	tetrafluorophthalic	PhMe	100	52:48
9	tetrafluorophthalic	Et ₂ O	100	85:15
10	tetrafluorophthalic	THF	100	>95:5
11	tetrachlorophthalic	THF	32	89:11
12	tetrachlorophthalic ^d	THF	74	80:20

^aX-ray crystal structure shown with 30% displacement ellipsoids (anion and hydrogen atoms omitted for clarity except for those on the ring). ^bConversion was determined by the ratio of product versus starting material by ¹H NMR analysis. ^cDiastereoselectivity determined by ¹H NMR analysis. ^dReaction run at reflux.

significant improvement in diastereoselectivity (entry 5). The commercially available and highly electron-deficient tetrafluorophthalic anhydride afforded 2-carboperoxy-3,4,5,6-tetrafluorobenzoic acid upon cleavage with hydrogen peroxide, whose structure features a more electrophilic peracid group tethered to a more acidic and strongly hydrogen bonding carboxylic acid group. This novel epoxidation reagent brought about high

Scheme 3. Controlling Face Selectivity in Epoxidation of Tetrahydropyridines Derived from Kinetic Protonation/Reduction Sequence by Tethering

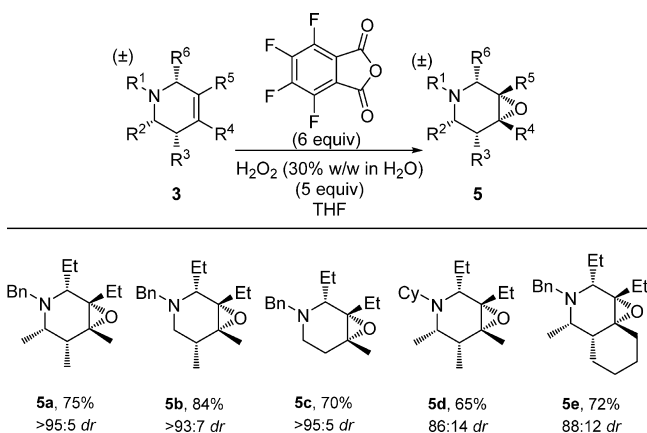


conversion in almost all solvents evaluated (entries 6–10). Moreover, a significant increase in diastereoselectivity was observed in ethereal solvents, particularly THF, which gave >95:5 dr (Table 2, entry 10). The slightly less electron-deficient tetrachlorophthalic anhydride required higher temperatures to bring about satisfactory conversion and resulted in lower diastereoselectivity (Table 2, entries 11 and 12). It is noteworthy that TFAA, which upon treatment with hydrogen peroxide also generates a highly electrophilic peracid and acidic carboxylic acid, was completely unreactive when THF was used as the solvent (entry 2). This result indicates that appropriate tethering of the peracid to the acid functionality is crucial for successful epoxidation. X-ray crystallographic analysis of protonated **5a** was performed to rigorously establish relative stereochemistry with the carboxy-substituted peracid enabling introduction of the epoxide oxygen on the significantly more sterically hindered face of the molecule (Table 2).

Scope for Stereoselective Epoxidation of 3. Encouraged by the high diastereoselectivity achieved in the epoxidation of **3a**, we next applied this novel epoxidation protocol to other tetrahydropyridines **3** obtained via a kinetic protonation/reduction sequence (Scheme 1a). High diastereoselectivities were observed for tetrahydropyridines with

differing degrees of substitution around the ring (**5a–c**) as well as branched alkyl substitution on the nitrogen (**5d**). Even the sterically hindered bicyclic epoxide **5e** was obtained with good diastereoselectivity (Table 3). The relative configuration of the major diastereoisomer was established by X-ray crystallography for epoxides **5a** and **5c** and assigned by analogy for the other products.

Table 3. Epoxidation of Tetrahydropyridines **3^a**



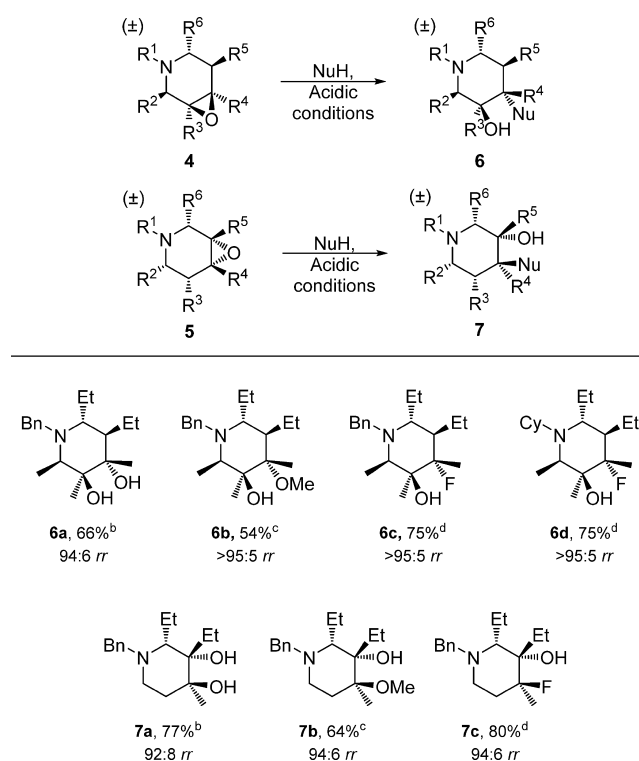
^aIsolated yields were determined by mass balance after purification by chromatography. Diastereoselectivities were determined by ^1H and ^{13}C NMR analysis. Relative configuration was rigorously assigned for **5a** and **5c** by X-ray structural analysis.

It is also noteworthy that the new epoxidation protocol was similarly effective for epoxidation of tetrahydropyridines **2** as demonstrated for **2a** (79% yield, >95:5 dr). Furthermore, for the epoxidation of tetrahydropyridine **2k**, the tetrafluorophthalic anhydride protocol resulted in a good yield and reasonable diastereoselectivity to give stereoisomer **4k** (Table 1), whereas the use of TFAA led to the formation of multiple side products.

Regioselective Opening of Epoxides 4 and 5. Epoxides **4** and **5** are versatile intermediates for the preparation of a variety of functionalized piperidine products via ring opening transformations. For epoxide **4**, the addition of fluoride, alcohol, and water under acidic conditions occurred with high regioselectivity to give highly substituted piperidinols **6** with adjacent tetrasubstituted carbons (Table 4). Specifically, addition of an OH group to give **6a** was achieved by heating in saturated aqueous sodium bisulfate,¹¹ a methoxy group was installed to give **6b** by heating in neat anhydrous methanol with benzenesulfonic acid, and fluorohydrins **6c** and **6d** were obtained by treatment with ethereal fluoroboric acid at room temperature.¹² The relative configuration and regiochemistry of the piperidinol products **6a** and **6b** were established by X-ray analysis. The observed high regioselectivity likely results from nucleophilic attack at the site distal from the deactivating protonated nitrogen of the piperidine ring.¹³

For epoxide **5c**, water, methanol and HF all cleanly added with high regio- and diastereoselectivity to give **7a**, **7b**, and **7c**, respectively. However, attempts to open more heavily substituted epoxides **5** derived from tetrahydropyridines **3** led to complex mixtures, presumably due to elimination and other side reactions.

Table 4. Nucleophilic Opening of Epoxides **4** and **5^a**



^aIsolated yields were determined by mass balance after purification by chromatography. Regioselectivities were determined by X-ray structural analysis for **6a** and **6b**, and for the remaining derivatives by ^1H , ^{13}C , and ^{19}F NMR analysis where applicable. ^b H_2O addition was achieved by heating at 70 °C in a $\text{CH}_2\text{Cl}_2/\text{sat. aq. NaHSO}_4$ mixture. ^cMeOH addition was performed at 80 °C in anhydrous MeOH with dry PhSO_3H (2 equiv). ^dHF addition was accomplished by treating with $\text{HBF}_4 \cdot \text{Et}_2\text{O}$ (2 equiv) at rt.

CONCLUSIONS

Highly diastereoselective epoxidations of tetrahydropyridines **2** and **3** have been demonstrated. Diastereoselective epoxidation of **2** was achieved by known methods of amino group protonation followed by epoxidation with a peracid. The diastereoselective epoxidation of **3** required the development of 2-carboperoxy-3,4,5,6-tetrafluorobenzoic acid, a new bifunctional epoxidation reagent that is prepared in situ from commercial materials, with the percarboxy acid group covalently tethered to the carboxylic acid functionality to enforce ammonium-directed epoxidation. A variety of nucleophiles were added to the epoxides **4** and **5** to provide piperidinols **6** and **7** with adjacent tetrasubstituted carbon stereocenters. Employing the methods reported here, high levels of substitution, functionalization, and stereocontrol can easily be achieved for the epoxide intermediates and the piperidinol products, with both compounds possessing valuable pharmaceutical potential.^{1,2,14}

EXPERIMENTAL SECTION

General Methods. Chromatography was performed on preparative thin-layer chromatography plates (1 mm SiO_2 , 20 × 20 cm). Molecular sieves were activated by heating to 280 °C in vacuo (ca. 0.1 Torr) for 6–12 h. For air-sensitive experiments, glassware was dried at 150 °C for at least 12 h and allowed to cool under an inert atmosphere. Experiments were set up inside a glovebox under a nitrogen atmosphere with oxygen and moisture levels not exceeding 1 ppm. Solvents for air-sensitive reactions were dried by passing through

activated alumina, degassed, and stored over 3 Å molecular sieves in a glovebox. Solvents of ACS reagent grade were used for workup and purification. Alkynes were distilled under a nitrogen atmosphere or in vacuo and stored in a glovebox prior to use. Liquid amines were purified according to procedures described in the literature¹⁶ and stored in a glovebox prior to use. $[\text{RhCl}(\text{coe})_2]_2$ was stored inside an N_2 -filled inert atmosphere glovebox at -25°C . The ligand, $\text{Me}_2\text{N}-\text{C}_6\text{H}_4-\text{PEt}_2$, was synthesized according to a previously published procedure^{7a} and stored inside a N_2 -filled inert atmosphere glovebox at -25°C . Stock solutions of the rhodium catalyst were made by dissolving $[\text{RhCl}(\text{coe})_2]_2$ (50.0 mg, 69.7 μmol) and $\text{Me}_2\text{N}-\text{C}_6\text{H}_4-\text{PEt}_2$ (30.0 mg, 143 μmol) in anhydrous PhMe until a total volume of 3.0 mL was reached. Stock solutions were used immediately and showed no difference in catalytic activity after being stored for months in a -25°C freezer inside a N_2 -filled glovebox. Methyl 4-methylpent-2-ynoate was prepared according to a literature procedure.¹⁷ All imines except (*E*)-1-cyclohexyl-*N*-((*E*)-3-methylpent-3-en-2-ylidene)-methanamine were prepared according to literature procedures.^{6,7a} With the exception of the tetrahydropyridines for which the preparations are described below, all tetrahydropyridines were prepared according to literature procedures.^{6,7b}

NMR characterization was performed on 400, 500, or 600 MHz instruments. Data are reported in the following format: chemical shift in ppm, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, dd = doublet of doublets, etc.), coupling constant *J* in hertz, integration, and interpretation. All spectra were referenced against residual solvent peaks (^1H : residual $\text{CHCl}_3 = 7.26$ ppm, ^{13}C : $\text{CDCl}_3 = 77.0$ ppm). For final products, d1 relaxation times of 15 s were used to record ^1H NMR spectra for accurate integration and quantitative determination of regio- and diastereoselectivities. IR spectra were obtained using FT-IR instruments in CH_2Cl_2 . Liquid chromatography–mass spectrometry (LC-MS) spectra were measured on an instrument equipped with dual atmospheric pressure chemical ionization (API)/electrospray ionization (ESI) and a photodiode array detector. High resolution mass spectra (ESI HRMS) were obtained on a time-of-flight (TOF) mass spectrometer.

(*E*)-1-Cyclohexyl-*N*-((*E*)-3-methylpent-3-en-2-ylidene)-methanamine (8). A 20 mL scintillation vial equipped with a stir bar was charged with 3-methylpent-3-en-2-one (363 mg, 3.70 mmol), cyclohexanemethylamine (423 mg, 3.74 mmol), dry THF (4 mL), and titanium(IV) ethoxide (4.0 mL, 18 mmol). The vial was capped and heated to 50°C for 2 h and then allowed to cool to rt. *N,N,N',N'*-tetrakis(2-hydroxyethyl)ethylenediamine (EDTE) (4.5 mL, 25 mmol) was added, and the mixture was heated to 55°C for 15 min until a clear yellow solution was observed. The mixture was cooled to rt and poured into a separatory funnel containing NH_4OH (20 mL) and brine (10 mL). The aqueous phase was extracted with EtOAc (30 mL), and the organic layer was washed with brine (20 mL), dried over MgSO_4 , filtered, and concentrated in vacuo. The residue was filtered through basic Al_2O_3 (ca. 3 cm in a Pasteur pipet, eluting with pentane), and the filtrate was concentrated in vacuo to give the desired imine (>98% *E*-isomer) as a slightly yellowish oil (480 mg, 2.48 mmol, 67%), which was stored at -25°C under an N_2 atmosphere in a glovebox. ^1H NMR (400 MHz, CDCl_3): δ 6.09–6.01 (m, 1 H), 3.15 (d, *J* = 6.7, 2 H), 1.90 (br s, 3 H), 1.84 (br s, 3 H), 1.82–1.58 (m, 7 H), 1.75 (d, *J* = 6.9, 3 H), 1.32–1.16 (m, 2 H), 1.00–0.87 (m, 2 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 166.4, 139.4, 126.6, 58.6, 39.6, 31.6, 26.8, 26.2, 14.4, 13.9, 12.8. IR (cm^{-1}): 3026, 2933, 2857, 1619, 1494, 1452, 1276, 1049, 731, 697. LC-MS: Exact mass calculated for $[\text{C}_{13}\text{H}_{23}\text{N} + \text{H}]^+$: 194.1909 *m/z*; found: 194.1920 *m/z*.

General Procedure for the Synthesis of Tetrahydropyridines 2d, 2k, and 2i from Imines and Alkynes. Inside a glovebox, the appropriate α,β -unsaturated imine (0.53 mmol), Rh stock solution (200 μL , 1.8 mol %), and 3-hexyne (138 μL , 1.21 mmol, 2.30 equiv) were combined in a 4 mL vial and transferred to a J. Young NMR tube equipped with a C_6D_6 capillary for locking and shimming. The vial was washed with PhMe (3 \times 10 drops), and the washings were transferred to the J. Young tube. More PhMe was added until a total volume of 0.6–0.7 mL was reached. The J. Young tube was capped and placed in an 80°C oil bath inside a fume hood for 3 h, at which point analysis by

^1H NMR indicated complete consumption of the imine. Inside a glovebox, a solution of diphenyl phosphate (320 mg, 1.28 mmol, 2.42 equiv) in anhydrous THF (0.3 mL) was added to the J. Young tube. The J. Young tube was capped and placed in a 50°C oil bath for 1 h, at which point ^1H NMR indicated full conversion into the C5-protonated iminium ion.

Inside a glovebox, a 20 mL scintillation vial equipped with a stir bar and a pierceable cap was charged with $\text{Na}(\text{AcO})_3\text{BH}$ (425 mg, 2.01 mmol, 3.80 equiv) and anhydrous THF (3 mL). The vial was capped and placed under a N_2 atmosphere at 0°C inside a fume hood. With vigorous stirring, the contents of the J. Young tube were added dropwise to the vial via a syringe and needle. The J. Young tube was subsequently washed with anhydrous THF (2 \times 0.4 mL), and the washings were transferred dropwise to the vial via the same syringe and needle. The reaction mixture was stirred at 0°C for a further 2 h and then allowed to warm to rt over 2 h.

The reaction was quenched with dH_2O (2 mL) and basified with 1 M NaOH until a pH of ca. 9 was reached. The aqueous phase was extracted with hexanes/EtOAc/ Et_3N (200:25:3, 3 \times 10 mL). The combined organic layers were filtered through SiO_2 (2.5 cm in a Pasteur pipet, deactivated with hexanes/EtOAc/ Et_3N (200:25:3)). The filtrate was concentrated under reduced pressure, and the crude product was purified by preparative TLC (SiO_2 , eluent hexanes/EtOAc/ Et_3N (200:25:3)) to afford tetrahydropyridines 2 and 3.

(2*R*,3*S*,6*R*)/(2*S*,3*R*,6*S*)-1-(Cyclohexylmethyl)-2,3-diethyl-4,5,6-trimethyl-1,2,3,6-tetrahydropyridine (2d). Employing the General Procedure using 8 (102 mg), 2d was obtained as a colorless oil (98.1 mg, 67%, dr > 20:1). ^1H NMR (500 MHz, CDCl_3): δ 2.65 (dd, *J* = 12.2, 6.0, 1 H), 2.54–2.48 (m, 1 H), 2.31 (dd, *J* = 13.2, 5.4, 1 H), 2.15 (dd, *J* = 13.2, 9.0, 1 H), 1.93–1.86 (m, 1 H), 1.75–1.33 (m, 9 H), 1.61 (br s, 3 H), 1.51 (br s, 3 H), 1.31–1.05 (m, 4 H), 1.01 (d, *J* = 6.3, 3 H), 0.90–0.72 (m, 2 H), 0.86 (dd, *J* = 7.3, 7.1, 3 H), 0.79 (dd, *J* = 7.5, 7.5, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 128.7, 126.9, 57.5, 57.0, 56.6, 45.5, 37.0, 32.1, 31.9, 27.1, 26.5, 26.3, 24.5, 19.8, 18.7, 17.0, 16.3, 12.5, 12.4. IR (cm^{-1}): 2956, 2930, 2853, 1451, 1376, 1338, 1307, 1260, 1198, 1181, 1155, 1113, 1094, 1072, 1051, 891, 842. LC-MS: Exact mass calculated for $[\text{C}_{19}\text{H}_{35}\text{N} + \text{H}]^+$: 278.2848 *m/z*; found: 278.2865 *m/z*.

(1*R*,3*R*,4*S*)/(1*S*,3*S*,4*R*)-2-Benzyl-3,4-diethyl-1-methyl-2,3,4,5,6,7-hexahydro-1*H*-cyclopenta[*c*]pyridine (2i). Employing the General Procedure using (*E*)-*N*-(1-(cyclopent-1-en-1-yl)ethylidene)-1-phenylmethanamine (78.2 mg), 2i was obtained as a yellowish oil (69.4 mg, 62%, dr > 20:1). ^1H NMR (500 MHz, CDCl_3): δ 7.44–7.39 (m, 2 H), 7.35–7.28 (m, 2 H), 7.26–7.20 (m, 1 H), 3.76 (d, *J* = 14.4, 1 H), 3.62 (d, *J* = 14.4, 1 H), 3.06 (dd, *J* = 12.8, 7.2, 1 H), 2.55–2.45 (m, 2 H), 2.43–2.34 (m, 1 H), 2.22–2.11 (m, 2 H), 1.92–1.80 (m, 3 H), 1.65–1.47 (m, 3 H), 1.37–1.26 (m, 1 H), 1.15 (d, *J* = 6.6, 3 H), 0.86 (dd, *J* = 7.4, 7.5, 3 H), 0.72 (dd, *J* = 7.5, 7.5, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 141.9, 136.9, 135.8, 128.3, 129.0, 126.2, 57.1, 53.1, 52.5, 40.0, 35.0, 33.6, 24.0, 22.5, 19.0, 18.1, 12.0, 11.4. IR (cm^{-1}): 2963, 2931, 2853, 1494, 1452, 1373, 1199, 1063, 1028, 967, 919, 756, 728, 697. LC-MS: Exact mass calculated for $[\text{C}_{20}\text{H}_{29}\text{N} + \text{H}]^+$: 284.2378 *m/z*; found: 284.2380 *m/z*.

(2*R*,3*S*,6*R*)/(2*S*,3*R*,6*S*)-1-Benzyl-2,3-diethyl-6-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (2k). Employing the General Procedure using (*E*)-1-phenyl-*N*-((*E*)-4-phenylbut-3-en-2-ylidene)methanamine (120 mg), 2k was obtained as a yellowish oil (117 mg, 72%, dr = 94:6). ^1H NMR (400 MHz, CDCl_3): δ 7.41–7.33 (m, 4 H), 7.33–7.27 (m, 4 H), 7.24–7.19 (m, 2 H), 5.76–5.74 (m, 1 H), 3.95 (d, *J* = 14.0, 1 H), 3.72 (d, *J* = 14.0, 1 H), 3.31–3.25 (m, 1 H), 2.56–2.51 (m, 1 H), 2.35–2.30 (m, 1 H), 1.78–1.69 (m, 1 H), 1.67–1.58 (m, 1 H), 1.41–1.26 (m, 2 H), 1.24 (d, *J* = 6.5, 3 H), 0.83 (dd, *J* = 7.4, 7.6, 3 H), 0.54 (dd, *J* = 7.4, 7.6, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (150 MHz, CDCl_3): δ 141.6, 141.1, 137.9, 128.8, 128.6, 128.3, 128.0, 126.7, 126.5, 125.8, 57.0, 53.7, 52.3, 41.8, 25.4, 21.1, 16.0, 12.5, 12.1. IR (cm^{-1}): 2963, 2931, 2871, 1494, 1453, 1374, 1067, 1029, 762, 724, 699, 642. LC-MS: Exact mass calculated for $[\text{C}_{23}\text{H}_{29}\text{N} + \text{H}]^+$: 320.2378 *m/z*; found: 320.2382 *m/z*.

(2*S*,3*S*,6*R*)/(2*R*,3*R*,6*S*)-1-Benzyl-5,6-diethyl-2,3,4-trimethyl-1,2,3,6-tetrahydropyridine (3b). Inside a glovebox, (*E*)-*N*-((*E*)-2-methylbut-

2-en-1-ylidene)-1-phenylmethanamine (82.2 mg, 0.473 mmol), Rh stock solution (150 μ L, 1.5 mol %), and 3-hexyne (118 μ L, 1.04 mmol, 2.20 equiv) were combined in a 4 mL vial and transferred to a J. Young NMR tube equipped with a C_6D_6 capillary for locking and shimming. The vial was washed with PhMe (3 \times 10 drops), and the washings were transferred to the J. Young tube. More PhMe was added until a total volume of 0.6–0.7 mL was reached. The J. Young tube was capped and placed in an 80 $^{\circ}$ C oil bath inside a fume hood for 2 h, at which point 1H NMR indicated complete consumption of the imine. Inside a glovebox, a 20 mL scintillation vial equipped with a stir bar and a pierceable cap was charged with $Na(AcO)_3BH$ (369 mg, 1.74 mmol, 3.68 equiv). The vial was capped and placed under a N_2 atmosphere at 0 $^{\circ}$ C inside a fume hood, and EtOH (3 mL) was added dropwise with stirring. The contents of the J. Young tube were transferred dropwise to the vial via a syringe and needle. The J. Young tube was subsequently washed with anhydrous THF (2 \times 0.4 mL), and the washings were transferred dropwise to the vial via the same syringe and needle. AcOH (1.5 mL) was added dropwise over 3 min, and the reaction mixture was stirred at 0 $^{\circ}$ C for a further 2 h and then allowed to warm to rt over 2 h. The reaction was quenched with deionized water (2 mL) and basified with 1 M NaOH until a pH of ca. 9 was reached. The aqueous phase was extracted with hexanes/EtOAc/Et $_3$ N (200:25:3, 3 \times 10 mL). The combined organic layers were filtered through SiO_2 (2.5 cm in a Pasteur pipet, deactivated with hexanes/EtOAc/Et $_3$ N (200:25:3)). The filtrate was concentrated under reduced pressure, and the crude product was purified by preparative TLC (SiO_2 , eluent hexanes/EtOAc/Et $_3$ N (200:25:3)) to afford **3b** as a yellowish oil (90.3 mg, 74%, dr = 88:12). 1H NMR (500 MHz, $CDCl_3$): δ 7.40–7.35 (m, 2 H), 7.33–7.28 (m, 2 H), 7.25–7.21 (m, 1 H), 3.75 (d, J = 13.5, 1 H), 3.54 (d, J = 13.5, 1 H), 2.68–2.63 (m, 1 H), 2.64 (dd, J = 13.0, 8.2, 1 H), 2.58–2.53 (m, 1 H), 2.20–2.05 (m, 2 H), 1.82–1.74 (m, 1 H), 1.67 (br s, 3 H), 1.64–1.56 (m, 1 H), 1.52–1.41 (m, 1 H), 0.95 (dd, J = 7.4, 7.6, 3 H), 0.95 (d, J = 6.9, 3 H), 0.90 (dd, J = 7.3, 7.3, 3 H). $^{13}C\{^1H\}$ NMR (125 MHz, $CDCl_3$): δ 140.7, 133.4, 129.2, 128.8, 128.0, 126.5, 62.3, 58.2, 52.2, 30.5, 24.5, 23.4, 17.5, 16.0, 13.1, 11.0. IR (cm^{-1}): 3026, 2958, 2929, 2870, 1494, 1452, 1364, 1271, 1119, 1073, 1041, 1028, 996, 733, 699. LC-MS: Exact mass calculated for $[C_{18}H_{27}N + H]^+$: 258.2222 m/z ; found: 258.2238 m/z .

General Procedure A for Preparation of Epoxides from Tetrahydropyridines 2 with mCPBA. A solution of the appropriate tetrahydropyridine **2** (0.4 mmol) in EtOAc (1 mL) was combined with a solution of CCl_3COOH (2 mmol) in EtOAc (1 mL) in an 8 mL vial, and the resulting mixture was stirred at room temperature for 5 min. A solution of mCPBA (0.9 mmol) in EtOAc (2 mL) was subsequently added, and the mixture was stirred for 2 h at room temperature. The reaction was quenched with sat. aq Na_2SO_3 (1 mL). Saturated aq $NaHCO_3$ was added until a pH of ca. 9 was reached. The mixture was extracted with hexanes/EtOAc/Et $_3$ N (200:25:3) (3 \times 5 mL), and the combined organic layers were filtered through silica gel (2.5 cm in a Pasteur pipet, deactivated with hexanes/EtOAc/Et $_3$ N (200:25:3)). The filtrate was concentrated under reduced pressure, and the crude product was purified by preparative TLC (SiO_2 , eluent hexanes/EtOAc/Et $_3$ N (200:25:3)) to afford epoxide **4**.

General Procedure B for Preparation of Epoxides from Tetrahydropyridines 2 with TFAA and H_2O_2 . At 0 $^{\circ}$ C, H_2O_2 (30% w/w in H_2O , 1.0 mmol) was added dropwise to a solution of trifluoroacetic anhydride (1.2 mmol) in CH_2Cl_2 (2 mL), and the mixture was stirred at 0 $^{\circ}$ C for 15 min. A solution of tetrahydropyridine **2** (0.21 mmol) in CH_2Cl_2 (1 mL) was then added, the ice–water bath was removed, and the mixture was allowed to warm to room temperature over 5 h. The reaction was quenched with sat. aq Na_2SO_3 (2 mL). Saturated aq $NaHCO_3$ was added until a pH of ca. 9 was reached. The mixture was extracted with hexanes/EtOAc/Et $_3$ N (200:25:3) (3 \times 5 mL), and the combined organic layers were filtered through silica gel (2.5 cm in a Pasteur pipet, deactivated with hexanes/EtOAc/Et $_3$ N (200:25:3)). The filtrate was concentrated under reduced pressure, and the crude product was purified by preparative TLC (SiO_2 , eluent hexanes/EtOAc/Et $_3$ N (200:25:3)) to afford epoxide **4**.

General Procedure C for Preparation of Epoxides from Tetrahydropyridines 3 and 2k with Tetrafluorophthalic Anhydride and H_2O_2 . At 0 $^{\circ}$ C, H_2O_2 (30% w/w in H_2O , 1.1 mmol) was added dropwise to a solution of tetrafluorophthalic anhydride (1.3 mmol) in THF (2 mL), and the mixture was stirred for 15 min. A solution of tetrahydropyridine **3** (0.22 mmol) in THF (1 mL) was then added, the ice–water bath was removed, and the mixture was allowed to warm to room temperature over 5 h. The reaction was quenched with sat. aq Na_2SO_3 (1 mL). Saturated aq $NaHCO_3$ was added until a pH of ca. 9 was reached. The mixture was extracted with hexanes/EtOAc/Et $_3$ N (200:25:3) (3 \times 5 mL), and the combined organic layers were filtered through silica gel (2.5 cm in a Pasteur pipet, deactivated with hexanes/EtOAc/Et $_3$ N (200:25:3)). The filtrate was concentrated under reduced pressure, and the crude product was purified by preparative TLC (SiO_2 , eluent hexanes/EtOAc/Et $_3$ N (200:25:3)) to afford epoxide **5** or **4k**.

Epoxide 4a. Employing General Procedure A using tetrahydropyridine **2a** (108 mg), **4a** was obtained as a colorless oil (106 mg, 93%, dr > 20:1). 1H NMR (500 MHz, $CDCl_3$): δ 7.34–7.30 (m, 2 H), 7.30–7.25 (m, 2 H), 7.22–7.18 (m, 1 H), 3.60 (d, J = 14.2, 1 H), 3.50 (d, J = 14.2, 1 H), 2.67 (dd, J = 6.4, 6.4, 1 H), 2.26–2.19 (m, 1 H), 1.70–1.48 (m, 2 H), 1.48–1.37 (m, 2 H), 1.33 (br s, 3 H), 1.30–1.20 (overlapping m, 1 H and br s, 3 H), 1.16 (d, J = 6.4, 3 H), 0.85 (dd, J = 7.4, 7.4, 3 H), 0.80 (dd, J = 7.5, 7.4, 3 H). $^{13}C\{^1H\}$ NMR (125 MHz, $CDCl_3$): δ 141.0, 128.3, 128.0, 126.5, 63.9, 62.7, 55.8, 54.5, 52.5, 41.2, 21.3, 21.1, 18.7, 18.6, 14.9, 12.3, 11.7. IR (cm^{-1}): 2960, 2930, 2873, 2805, 1494, 1453, 1379, 1149, 1140, 1112, 1078, 1061, 1037, 1027, 897, 869, 855, 834, 805. LC-MS: Exact mass calculated for $[C_{19}H_{29}NO + H]^+$: 288.2327 m/z ; found: 288.2354 m/z .

Epoxide 4b. Employing General Procedure A using tetrahydropyridine **2b** (74.5 mg), **4b** was obtained as a yellowish oil (59.6 mg, 76%, dr = 84:16). 1H NMR (500 MHz, $CDCl_3$): δ 7.24–7.19 (m, 2 H), 6.88–6.83 (m, 2 H), 6.75–6.70 (m, 1 H), 4.01–3.97 (m, 1 H), 3.73–3.65 (m, 1 H), 2.14–2.07 (m, 1 H), 2.05–1.96 (m, 1 H), 1.89–1.79 (m, 1 H), 1.73–1.64 (m, 1 H), 1.49–1.40 (m, 1 H), 1.32 (br s, 3 H), 1.13 (d, J = 7.1, 3 H), 1.09 (d, J = 7.1, 3 H), 1.04 (dd, J = 7.6, 7.0, 3 H), 1.01 (dd, J = 7.4, 7.6, 3 H). $^{13}C\{^1H\}$ NMR (125 MHz, $CDCl_3$): δ 149.6, 129.1, 117.6, 115.9, 68.5, 64.8, 55.9, 52.2, 37.1, 26.7, 25.4, 16.7, 15.3, 13.1, 12.9, 10.3. IR (cm^{-1}): 2964, 2927, 2874, 1596, 1499, 1458, 1377, 1306, 1280, 1118, 1071, 1039, 990, 974, 890, 861, 812. LC-MS: Exact mass calculated for $[C_{18}H_{27}NO + H]^+$: 274.2171 m/z ; found: 274.2163 m/z .

Epoxide 4c. Employing General Procedure A using tetrahydropyridine **2c** (120 mg), **4c** was obtained as a yellowish oil (117 mg, 92%, dr > 20:1). 1H NMR (500 MHz, $CDCl_3$): δ 2.99 (dd, J = 12.8, 6.5, 1 H), 2.63–2.56 (m, 1 H), 2.38–2.33 (m, 1 H), 1.79–1.69 (m, 3 H), 1.64–1.50 (m, 4 H), 1.48–1.28 (m, 5 H), 1.26 (s, 3 H), 1.26–1.21 (m, 1 H), 1.23 (s, 3 H), 1.09 (d, J = 6.4, 3 H), 0.93 (dd, J = 7.4, 7.6, 3 H), 0.84 (dd, J = 7.4, 7.4, 3 H). $^{13}C\{^1H\}$ NMR (125 MHz, $CDCl_3$): δ 64.2, 63.6, 55.1, 54.2, 50.3, 42.4, 33.3, 31.1, 26.9, 26.8, 26.4, 23.1, 21.7, 21.5, 17.6, 14.8, 12.5, 11.7. IR (cm^{-1}): 2963, 2928, 1493, 1449, 1373, 1261, 1201, 1192, 1118, 1078, 1027, 892, 599. LC-MS: Exact mass calculated for $[C_{18}H_{33}NO + H]^+$: 280.2640 m/z ; found: 280.2675 m/z .

Epoxide 4d. Employing General Procedure A using tetrahydropyridine **2d** (84.1 mg), **4d** was obtained as a colorless oil (72.1 mg, 92%, dr > 20:1). 1H NMR (500 MHz, $CDCl_3$): δ 2.64 (dd, J = 12.6, 6.2, 1 H), 2.18–2.14 (m, 1 H), 2.13–2.04 (m, 2 H), 1.80–1.58 (m, 6 H), 1.51–1.38 (m, 3 H), 1.28 (s, 3 H), 1.20 (s, 3 H), 1.27–1.08 (m, 5 H), 1.06 (d, J = 6.4, 3 H), 0.93 (dd, J = 7.4, 7.2, 3 H), 0.84 (dd, J = 7.4, 7.4, 3 H), 0.79–0.67 (m, 2 H). $^{13}C\{^1H\}$ NMR (125 MHz, $CDCl_3$): δ 63.3, 63.0, 58.3, 56.2, 55.0, 41.3, 37.6, 31.8, 31.8, 27.0, 26.3, 26.3, 21.9, 21.6, 18.4, 18.2, 15.2, 12.6, 12.1. IR (cm^{-1}): 2958, 2922, 2873, 2851, 1449, 1379, 1163, 1137, 1113, 1082, 1059, 891, 870, 856. LC-MS: Exact mass calculated for $[C_{19}H_{35}NO + H]^+$: 294.2797 m/z ; found: 294.2791 m/z .

Epoxide 4e. Employing General Procedure A using tetrahydropyridine **2e** (56.9 mg), **4e** was obtained as a yellowish oil (49.9 mg, 83%, dr = 94:6). 1H NMR (500 MHz, $CDCl_3$): δ 7.39–7.35 (m, 2 H), 7.30–7.26 (m, 2 H), 7.21–7.17 (m, 1 H), 3.99 (d, J = 15.4, 1 H), 3.72 (d, J = 15.4, 1 H), 3.12–2.99 (m, 1 H), 2.21 (d, J = 5.4, 1 H), 1.93–

1.86 (m, 1 H), 1.38 (br s, 3 H), 1.21 (d, $J = 6.9$, 3 H), 1.18 (s, 3 H), 1.11 (d, $J = 6.8$, 3 H), 0.99 (s, 9 H). The $^{13}\text{C}\{^1\text{H}\}$ NMR for epoxide **4e** was not informative due to slow rotation at room temperature on the ^{13}C NMR time scale. IR (cm^{-1}): 2958, 2929, 2873, 1494, 1453, 1367, 1171, 1149, 1124, 1061, 1028, 990, 971, 905, 861. LC-MS: Exact mass calculated for $[\text{C}_{20}\text{H}_{31}\text{NO} + \text{H}]^+$: 302.2484 m/z ; found: 302.2479 m/z .

Epoxide 4f. Employing General Procedure B using tetrahydropyridine **2f** (66.3 mg), **4f** was obtained as a yellowish oil (48.8 mg, 70%, $\text{dr} > 20:1$). ^1H NMR (500 MHz, CDCl_3): δ 7.35–7.28 (m, 4 H), 7.25–7.21 (m, 1 H), 3.85 (d, $J = 14.6$, 1 H), 3.74 (s, 3 H, ester CH_3), 3.25 (d, $J = 14.6$, 1 H), 2.85 (d, $J = 1.2$, 1 H), 2.84–2.83 (m, 1 H), 2.75 (dd, $J = 13.7$, 6.9, 1 H), 1.83–1.74 (m, 1 H), 1.40 (s, 3 H), 1.20 (br s, 3 H), 1.12 (d, $J = 6.8$, 3 H), 1.02 (d, $J = 6.8$, 3 H), 0.89 (d, $J = 6.6$, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 173.6, 140.1, 128.3, 127.8, 126.8, 62.1, 61.4, 56.4, 54.3, 51.8, 51.7, 46.9, 29.8, 21.2, 20.4, 20.1, 17.6, 13.9. IR (cm^{-1}): 2951, 1739, 1494, 1453, 1434, 1381, 1367, 1249, 1189, 1139, 1101, 1062, 1026, 973, 948, 920, 851. LC-MS: Exact mass calculated for $[\text{C}_{20}\text{H}_{29}\text{NO}_3 + \text{H}]^+$: 332.2226 m/z ; found: 332.2210 m/z .

Epoxide 4g. Employing General Procedure A using tetrahydropyridine **2g** (63.8 mg), **4g** was obtained as a yellowish oil (53.5 mg, 79%, $\text{dr} > 20:1$). ^1H NMR (500 MHz, CDCl_3): δ 7.34–7.25 (m, 4 H), 7.23–7.18 (m, 1 H), 3.63 (d, $J = 13.5$, 1 H), 3.59 (d, $J = 13.5$, 1 H), 2.83 (d, $J = 13.3$, 1 H), 2.57 (d, $J = 13.3$, 1 H), 2.16–2.11 (m, 1 H), 1.74–1.56 (m, 2 H), 1.55–1.44 (m, 1 H), 1.40–1.35 (m, 1 H), 1.32 (s, 3 H), 1.24 (s, 3 H), 1.12–1.01 (m, 1 H), 0.81 (dd, $J = 7.3$, 7.4, 3 H), 0.81 (dd, $J = 7.5$, 7.4, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 139.5, 128.6, 128.1, 126.7, 61.8, 60.2, 58.4, 58.2, 51.9, 42.7, 21.9, 21.7, 17.6, 17.4, 12.5, 11.6. IR (cm^{-1}): 2957, 2930, 2870, 1494, 1452, 1364, 1280, 1120, 1074, 1041, 1028, 996, 732. LC-MS: Exact mass calculated for $[\text{C}_{18}\text{H}_{27}\text{NO} + \text{H}]^+$: 274.2171 m/z ; found: 274.2175 m/z .

Epoxide 4h. Employing General Procedure B using tetrahydropyridine **2h** (95.7 mg), **4h** was obtained as a yellowish oil (90.8 mg, 90%, $\text{dr} > 20:1$). ^1H NMR (500 MHz, CDCl_3): δ 7.35–7.30 (m, 2 H), 7.30–7.24 (m, 2 H), 7.22–7.17 (m, 1 H), 3.63 (d, $J = 14.3$, 1 H), 3.49 (d, $J = 14.3$, 1 H), 2.70 (dd, $J = 12.8$, 6.4, 1 H), 2.29–2.23 (m, 1 H), 2.11–2.04 (m, 1 H), 1.90–1.82 (m, 1 H), 1.67–1.37 (m, 8 H), 1.37–1.20 (m, 3 H), 1.11 (d, $J = 6.4$, 3 H), 0.86 (dd, $J = 7.4$, 7.4, 3 H), 0.78 (dd, $J = 7.5$, 7.2, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 141.1, 128.6, 128.0, 126.4, 63.8, 62.4, 55.8, 55.1, 52.3, 41.4, 31.9, 29.7, 21.1, 20.8, 20.4, 18.8, 14.5, 12.0, 11.8. IR (cm^{-1}): 2958, 2932, 2873, 1494, 1453, 1374, 1178, 1159, 1103, 1068, 1027, 956, 880, 869, 849, 832, 815. LC-MS: Exact mass calculated for $[\text{C}_{21}\text{H}_{31}\text{NO} + \text{H}]^+$: 314.2484 m/z ; found: 314.2451 m/z .

Epoxide 4i. Employing General Procedure B using tetrahydropyridine **2i** (72.0 mg), **4i** was obtained as a yellowish oil (63.9 mg, 84%, $\text{dr} > 20:1$). ^1H NMR (500 MHz, CDCl_3): δ 7.36–7.32 (m, 2 H), 7.31–7.26 (m, 2 H), 7.23–7.18 (m, 1 H), 3.57 (d, $J = 14.3$, 1 H), 3.53 (d, $J = 14.3$, 1 H), 2.92 (dd, $J = 13.1$, 6.6, 1 H), 2.31–2.25 (m, 1 H), 2.17–2.12 (m, 1 H), 2.01–1.94 (m, 1 H), 1.74–1.25 (m, 9 H), 1.17 (d, $J = 6.5$, 3 H), 0.90 (dd, $J = 7.2$, 7.4, 3 H), 0.89 (dd, $J = 7.0$, 7.4, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 141.2, 128.2, 128.1, 126.5, 69.5, 68.3, 56.7, 51.8, 51.7, 38.4, 33.0, 31.0, 22.5, 19.9, 19.3, 15.8, 12.1, 11.9. IR (cm^{-1}): 2959, 2931, 2873, 1454, 1368, 1175, 1130, 1106, 1074, 1027, 972, 922, 890, 840. LC-MS: Exact mass calculated for $[\text{C}_{20}\text{H}_{29}\text{NO} + \text{H}]^+$: 300.2327 m/z ; found: 300.2338 m/z .

Epoxide 4j. Employing General Procedure B using tetrahydropyridine **2j** (89.4 mg), **4j** was obtained as a yellowish oil (80.6 mg, 86%, $\text{dr} = 91:9$). ^1H NMR (500 MHz, CDCl_3): δ 7.44–7.18 (m, 10 H), 3.74 (m, 2 H), 2.82 (dd, $J = 13.0$, 6.6, 1 H), 2.59–2.51 (m, 1 H), 2.06–2.00 (m, 1 H), 1.65–1.50 (m, 3 H), 1.42–1.32 (m, 1 H), 1.22 (d, $J = 6.5$, 3 H), 1.01 (dd, $J = 7.3$, 7.3, 3 H), 0.86 (s, 3 H), 0.66 (dd, $J = 7.6$, 7.6, 3 H). The $^{13}\text{C}\{^1\text{H}\}$ NMR for epoxide **4j** was not informative due to slow rotation at room temperature on the ^{13}C NMR time scale. IR (cm^{-1}): 3060, 3026, 2961, 2931, 2874, 1658, 1603, 1494, 1446, 1375, 1200, 1155, 1122, 1090, 1069, 1051, 1027, 988, 956, 920, 898, 883, 840. LC-MS: Exact mass calculated for $[\text{C}_{24}\text{H}_{31}\text{NO} + \text{H}]^+$: 350.2484 m/z ; found: 350.2478 m/z .

Epoxide 4k. Employing General Procedure C using tetrahydropyridine **2k** (52.5 mg), **4k** was obtained as a yellowish oil (30.3 mg, 55%, $\text{dr} = 86:14$). ^1H NMR (500 MHz, CDCl_3): δ 7.44–7.12 (m, 10 H), 3.79 (d, $J = 14.4$, 1 H), 3.58 (d, $J = 14.4$, 1 H), 3.06–3.00 (m, 1 H), 2.86–2.83 (m, 1 H), 2.42–2.33 (m, 1 H), 2.16–2.11 (m, 1 H), 1.74–1.59 (m, 2 H), 1.59–1.45 (m, 2 H), 1.45–1.35 (m, 1 H), 1.26 (d, $J = 6.5$, 3 H), 0.97 (dd, $J = 7.3$, 7.6, 3 H), 0.63 (dd, $J = 7.5$, 7.5, 3 H). The $^{13}\text{C}\{^1\text{H}\}$ NMR for epoxide **4k** was not informative due to slow rotation at room temperature on the ^{13}C NMR time scale. IR (cm^{-1}): 3027, 2962, 2932, 2872, 1659, 1604, 1494, 1375, 1200, 1155, 1122, 1090, 1068, 1052, 1027, 988, 898. LC-MS: Exact mass calculated for $[\text{C}_{23}\text{H}_{29}\text{NO} + \text{H}]^+$: 336.2327 m/z ; found: 336.2346 m/z .

Epoxide 5a. Employing General Procedure C using tetrahydropyridine **3a** (70.7 mg), **5a** was obtained as a colorless oil (56.2 mg, 75%, $\text{dr} > 20:1$). ^1H NMR (500 MHz, CDCl_3): δ 7.38–7.34 (m, 2 H), 7.31–7.27 (m, 2 H), 7.24–7.20 (m, 1 H), 3.73 (d, $J = 13.9$, 1 H), 3.66 (d, $J = 13.9$, 1 H), 2.76–2.69 (m, 1 H), 2.65 (dd, $J = 8.6$, 4.9, 1 H), 2.17–2.11 (m, 1 H), 1.83–1.74 (m, 1 H), 1.76–1.66 (m, 1 H), 1.61–1.52 (m, 1 H), 1.36–1.30 (m, 1 H), 1.34 (br s, 3 H), 1.13 (d, $J = 7.4$, 3 H), 1.00 (d, $J = 7.1$, 3 H), 0.99 (dd, $J = 7.6$, 7.6, 3 H), 0.88 (dd, $J = 7.5$, 7.5, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 140.9, 128.6, 128.0, 126.7, 67.9, 64.6, 59.8, 59.2, 56.6, 32.6, 26.4, 23.7, 16.9, 16.4, 12.8, 12.5, 10.1. IR (cm^{-1}): 2961, 2931, 2874, 1726, 1461, 1379, 1271, 1122, 1071, 1041, 1027, 957, 895, 874, 842. LC-MS: Exact mass calculated for $[\text{C}_{19}\text{H}_{29}\text{NO} + \text{H}]^+$: 288.2327 m/z ; found: 288.2303 m/z .

Epoxide 5b. Employing General Procedure C using tetrahydropyridine **3b** (55.2 mg), **5b** was obtained as a yellowish oil (49.3 mg, 84%, $\text{dr} = 93:7$). ^1H NMR (500 MHz, CDCl_3): δ 7.38–7.33 (m, 2 H), 7.33–7.26 (m, 2 H), 7.26–7.19 (m, 1 H), 3.68 (d, $J = 13.5$, 1 H), 3.56 (d, $J = 13.5$, 1 H), 2.53–2.48 (m, 1 H), 2.42 (dd, $J = 14.0$, 11.4, 1 H), 2.35–2.29 (m, 1 H), 2.07–1.99 (m, 1 H), 1.72–1.63 (m, 1 H), 1.61–1.51 (m, 1 H), 1.46–1.39 (m, 1 H), 1.39–1.30 (m, 1 H), 1.37 (s, 3 H), 0.98 (dd, $J = 7.6$, 7.6, 3 H), 0.88 (d, $J = 6.7$, 3 H), 0.85 (dd, $J = 7.6$, 7.4, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 140.2, 129.3, 128.5, 127.3, 67.3, 63.7, 58.8, 58.6, 48.6, 28.7, 27.1, 21.1, 16.6, 13.8, 12.3, 10.2. IR (cm^{-1}): 2963, 2931, 2872, 1493, 1453, 1374, 1281, 1121, 1073, 1028, 761, 725, 643. LC-MS: Exact mass calculated for $[\text{C}_{18}\text{H}_{27}\text{NO} + \text{H}]^+$: 274.2171 m/z ; found: 274.2167 m/z .

Epoxide 5c. Employing General Procedure C using tetrahydropyridine **3c** (45.9 mg), **5c** was obtained as a yellowish oil (34.2 mg, 70%, $\text{dr} > 20:1$). ^1H NMR (500 MHz, CDCl_3): δ 7.38–7.34 (m, 2 H), 7.32–7.28 (m, 2 H), 7.26–7.21 (m, 1 H), 3.67 (d, $J = 13.4$, 1 H), 3.56 (d, $J = 13.4$, 1 H), 2.89–2.82 (m, 1 H), 2.52–2.47 (m, 1 H), 2.43–2.38 (m, 1 H), 2.03–1.94 (m, 1 H), 1.72–1.64 (m, 1 H), 1.64–1.54 (m, 1 H), 1.53–1.47 (m, 1 H), 1.47–1.35 (m, 2 H), 1.40 (br s, 3 H), 0.99 (dd, $J = 7.6$, 7.6, 3 H), 0.86 (dd, $J = 7.4$, 7.4, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 140.0, 128.9, 128.0, 126.9, 66.1, 60.5, 58.6, 57.6, 40.3, 26.3, 26.0, 20.8, 19.2, 11.9, 9.9. IR (cm^{-1}): 2962, 2930, 2874, 2805, 1454, 1376, 1138, 1109, 1082, 1074, 1058, 1009, 976, 902, 882, 856, 833. LC-MS: Exact mass calculated for $[\text{C}_{17}\text{H}_{25}\text{NO} + \text{H}]^+$: 260.2014 m/z ; found: 260.1970 m/z .

Epoxide 5d. Employing General Procedure C using tetrahydropyridine **3d** (58.0 mg), **5d** was obtained as a yellowish oil (40.0 mg, 65% with respect to the tetrahydropyridine, $\text{dr} = 86:14$). ^1H NMR (500 MHz, CDCl_3): δ 2.91–2.78 (m, 2 H), 2.38–2.27 (m, 1 H), 1.99–1.64 (m, 6 H), 1.63–1.49 (m, 3 H), 1.42–1.10 (m, 6 H), 1.27 (s, 3 H), 1.07 (d, $J = 7.4$, 3 H), 0.99 (dd, $J = 7.9$, 7.4, 3 H), 0.97 (d, $J = 7.1$, 3 H), 0.96 (dd, $J = 7.4$, 7.5, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 68.9, 64.9, 64.9, 57.6, 51.3, 36.2, 32.9, 32.8, 26.9, 26.5, 26.4, 26.2, 25.1, 18.2, 16.5, 14.1, 11.9, 10.6. IR (cm^{-1}): 2962, 2930, 1494, 1461, 1451, 1375, 1261, 1199, 1150, 1115, 1078, 1027, 944, 892, 696. LC-MS: Exact mass calculated for $[\text{C}_{18}\text{H}_{34}\text{NO} + \text{H}]^+$: 280.2640 m/z ; found: 280.2666 m/z .

Epoxide 5e. Employing General Procedure C using tetrahydropyridine **3e** (51.7 mg), **5e** was obtained as a yellowish oil (39.2 mg, 72% with respect to the tetrahydropyridine, $\text{dr} = 88:12$). ^1H NMR (500 MHz, CDCl_3): δ 7.40–7.14 (m, 5 H), 3.74 (d, $J = 13.6$, 1 H), 3.64 (d, $J = 13.6$, 1 H), 2.76–2.69 (m, 1 H), 2.64–2.60 (m, 1 H), 2.11–2.06 (m, 1 H), 1.90–1.31 (m, 12 H), 1.16 (d, $J = 7.3$, 3 H), 0.96 (dd, $J = 7.6$, 7.6, 3 H), 0.86 (dd, $J = 7.3$, 7.4, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz,

CDCl_3): δ 140.7, 128.8, 128.0, 126.8, 67.8, 65.7, 60.5, 58.8, 55.4, 35.2, 30.2, 27.7, 26.4, 25.9, 24.3, 23.5, 17.0, 12.4, 10.1. IR (cm^{-1}): 2956, 2933, 2872, 1494, 1454, 1372, 1199, 1179, 1061, 1027, 943, 884, 869, 849, 832, 815. LC-MS: Exact mass calculated for $[\text{C}_{21}\text{H}_{31}\text{NO} + \text{H}]^+$: 314.2484 m/z ; found: 314.2460 m/z .

General Procedure for Water Opening of Epoxides 4 and 5.

In a 4 mL vial equipped with a stir bar, a solution of epoxide 4 or 5 (0.2 mmol) in CH_2Cl_2 (0.2 mL) was added to sat. aq NaHSO_4 (2 mL). The vial was capped and heated to 70 °C for 12 h with stirring. The reaction mixture was allowed to cool to room temperature, and sat. aq NaHCO_3 added until a pH of ca. 9 was reached. The resultant mixture was extracted with hexanes/EtOAc/Et₃N (200:25:3 v:v:v) (3 × 5 mL), and the combined organic layers were filtered through silica gel (2.5 cm in a Pasteur pipet, deactivated with hexanes/EtOAc/Et₃N (200:25:3)). The filtrate was concentrated under reduced pressure, and the crude product was purified by preparative TLC (SiO_2 , eluent hexanes/EtOAc/Et₃N (200:25:3)) to afford product 6 or 7.

Product 6a. Employing the General Procedure using epoxide 4a (51.5 mg), 6a was obtained as an off-white solid (mp 110–112 °C, 36.2 mg, 66%, $rr = 94:6$). ^1H NMR (500 MHz, CDCl_3): δ 7.32–7.25 (m, 4 H), 7.24–7.19 (m, 1 H), 3.99 (d, $J = 13.1$, 1 H), 3.51 (d, $J = 13.1$, 1 H), 3.35 (dd, $J = 13.6$, 6.8, 1 H), 3.35 (br s, 1 H), 2.27 (dd, $J = 10.8$, 2.6, 1 H), 2.12–2.01 (m, 1 H), 1.71–1.60 (m, 1 H), 1.57–1.48 (m, 1 H), 1.27 (br s, 3 H), 1.19–1.17 (m, 1 H), 1.12–1.09 (m, 1 H), 1.07 (br s, 3 H), 1.04 (d, $J = 6.8$, 3 H), 0.74 (dd, $J = 7.4$, 7.4, 3 H), 0.52 (dd, $J = 8.2$, 7.4, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 140.7, 128.6, 128.2, 126.8, 76.7, 74.1, 59.6, 53.3, 53.3, 49.2, 25.4, 24.3, 18.2, 17.2, 14.4, 12.6, 11.4. IR (cm^{-1}): 3435, 2967, 2873, 1453, 1382, 1279, 1190, 1175, 1106, 1081, 1028, 997, 956, 921, 900, 837. LC-MS: Exact mass calculated for $[\text{C}_{19}\text{H}_{31}\text{NO}_2 + \text{H}]^+$: 306.2433 m/z ; found: 306.2435 m/z .

Product 7a. Employing the General Procedure using epoxide 5c (48.4 mg), 7a was obtained as a colorless oil (39.9 mg, 77%, $rr = 92:8$). ^1H NMR (500 MHz, CDCl_3): δ 7.33–7.28 (m, 4 H), 7.26–7.22 (m, 1 H), 4.14 (d, $J = 13.2$, 1 H), 3.37 (br s, 1 H), 3.07 (d, $J = 13.2$, 1 H), 2.68 (dd, $J = 3.7$, 3.7, 1 H), 3.58–3.52 (m, 1 H), 2.30–2.22 (m, 1 H), 1.97–1.71 (m, 4 H), 1.68–1.57 (m, 2 H), 1.32–1.27 (br s, 1 H), 1.24 (br s, 3 H), 1.09 (dd, $J = 7.5$, 7.6, 3 H), 1.03 (dd, $J = 7.8$, 7.9, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 140.0, 128.5, 128.3, 126.9, 75.6, 74.4, 63.9, 57.4, 47.5, 36.4, 25.9, 24.9, 20.3, 12.3, 9.6. IR (cm^{-1}): 3472, 2965, 2939, 2879, 2825, 1495, 1452, 1386, 1366, 1306, 1268, 1169, 1106, 1084, 1044, 1028, 1007, 922, 867, 840, 828, 809. LC-MS: Exact mass calculated for $[\text{C}_{17}\text{H}_{27}\text{NO}_2 + \text{H}]^+$: 278.2120 m/z ; found: 278.2101 m/z .

General Procedure for Methoxide Opening of Epoxides 4 and 5. In an oven-dried 4 mL vial equipped with a stir bar, epoxide 4 or 5 (0.2 mmol) and anhydrous benzenesulfonic acid (0.4 mmol) were dissolved in anhydrous MeOH (1.5 mL, stored over 3 Å molecular sieves for 2 days). The vial was capped, sealed with Parafilm, and heated to 80 °C for 7 h with stirring. The reaction mixture was allowed to cool to room temperature, and the volatiles were removed under a stream of N_2 . Saturated aq NaHCO_3 was then added until a pH of ca. 9 was reached. The resultant mixture was extracted with hexanes/EtOAc/Et₃N (200:25:3 v:v:v) (3 × 5 mL), and the combined organic layers were filtered through silica gel (2.5 cm in a Pasteur pipet, deactivated with hexanes/EtOAc/Et₃N (200:25:3)). The filtrate was concentrated under reduced pressure, and the crude product was purified by preparative TLC (SiO_2 , eluent hexanes/EtOAc/Et₃N (200:25:3)) to afford product 6 or 7.

Product 6b. Employing the General Procedure using epoxide 4a (49.2 mg), 6b was obtained as a yellowish solid (mp 80–82 °C, 29.6 mg, 54%, $rr > 20:1$). ^1H NMR (500 MHz, CDCl_3): δ 7.31–7.24 (overlapping m, 4 H), 7.23–7.19 (m, 1 H), 3.97 (d, $J = 13.1$, 1 H), 3.57 (br s, 1 H), 3.48 (d, $J = 13.1$, 1 H), 3.34 (dd, $J = 13.7$, 7.0, 1 H), 3.08 (s, 3 H), 2.17 (dd, $J = 11.0$, 2.0, 1 H), 1.93–1.83 (m, 1 H), 1.68–1.59 (m, 1 H), 1.42–1.33 (m, 2 H), 1.19 (br s, 3 H), 1.20–1.12 (m, 1 H), 1.01 (d, $J = 6.9$, 3 H), 1.00 (br s, 3 H), 0.74 (dd, $J = 7.4$, 7.5, 3 H), 0.51 (dd, $J = 7.3$, 7.4, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 140.9, 128.6, 128.2, 126.7, 80.4, 74.6, 59.8, 53.3, 53.1, 48.4, 40.1, 23.8, 19.0, 18.2, 16.5, 14.4, 12.8, 11.8. IR (cm^{-1}): 3435, 2958, 2874, 2828,

1495, 1454, 1381, 1365, 1288, 1267, 1185, 1147, 1129, 1092, 1046, 1028, 997, 920, 896, 854. LC-MS: Exact mass calculated for $[\text{C}_{20}\text{H}_{33}\text{NO}_2 + \text{H}]^+$: 320.2590 m/z ; found: 320.2570 m/z .

Product 7b. Employing the General Procedure using epoxide 5c (43.3 mg), 7b was obtained as a yellowish oil (31.1 mg, 54%, $rr = 94:6$). ^1H NMR (500 MHz, CDCl_3): δ 7.33–7.28 (m, 4 H), 7.26–7.20 (m, 1 H), 4.16 (d, $J = 13.1$, 1 H), 3.52 (br s, 1 H), 3.11 (s, 3 H), 3.00 (d, $J = 13.1$, 1 H), 2.72 (dd, $J = 3.6$, 3.6, 1 H), 2.47–2.40 (m, 1 H), 2.14–2.04 (m, 1 H), 1.96–1.86 (m, 1 H), 1.74 (dd, $J = 15.7$, 7.8, 2 H), 1.64–1.51 (m, 3 H), 1.13 (br s, 3 H), 1.09 (dd, $J = 7.6$, 7.7, 3 H), 0.99 (dd, $J = 7.7$, 7.9, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 140.3, 128.5, 128.3, 126.8, 78.5, 75.6, 63.6, 57.4, 48.1, 47.6, 28.8, 26.1, 20.6, 18.4, 12.3, 9.7. IR (cm^{-1}): 2967, 2940, 2879, 2825, 1452, 1387, 1366, 1306, 1242, 1176, 1160, 1121, 1095, 1066, 1028, 1007, 967, 909, 889, 867, 839, 828. LC-MS: Exact mass calculated for $[\text{C}_{18}\text{H}_{29}\text{NO}_2 + \text{H}]^+$: 292.2277 m/z ; found: 292.2231 m/z .

General Procedure for Fluoride Opening of Epoxides 4 and 5.

In a 4 mL vial equipped with a stir bar, $\text{HBF}_4 \cdot \text{Et}_2\text{O}$ ("51–57% in Et_2O ", ca. 0.4 mmol) was added to a solution of epoxide 4 or 5 (0.2 mmol) in CH_2Cl_2 (1 mL). The vial was capped, and the mixture was stirred at room temperature for 10 min. Saturated aq NaHCO_3 was then added until a pH of ca. 9 was reached. The resultant mixture was extracted with hexanes/EtOAc/Et₃N (200:25:3 v:v:v) (3 × 5 mL), and the combined organic layers were filtered through silica gel (2.5 cm in a Pasteur pipet, deactivated with hexanes/EtOAc/Et₃N (200:25:3)). The filtrate was concentrated under reduced pressure, and the crude product was purified by preparative TLC (SiO_2 , eluent hexanes/EtOAc/Et₃N (200:25:3)) to afford product 6 or 7.

Product 6c. Employing the General Procedure using epoxide 4a (63.5 mg), 6c was obtained as an orange oil (50.7 mg, 75%, $rr > 20:1$). ^1H NMR (400 MHz, CDCl_3): δ 7.32–7.24 (m, 4 H), 7.24–7.20 (m, 1 H), 3.99 (d, $J = 13.1$, 1 H), 3.51 (d, $J = 13.1$, 1 H), 3.33–3.24 (overlapping br s and m, 2 H), 2.26 (dd, $J = 11.1$, 2.5, 1 H), 1.89–1.78 (m, 1 H), 1.68–1.59 (m, 1 H), 1.51–1.38 (m, 2 H), 1.38 (d, $J = 23.5$, 3 H), 1.20–1.14 (m, 1 H), 1.10 (d, $J = 2.0$, 3 H), 1.07 (d, $J = 6.8$, 3 H), 0.73 (dd, $J = 7.4$, 7.4, 3 H), 0.54 (dd, $J = 7.2$, 7.3, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 140.5, 128.6, 128.3, 126.9, 99.8 (d, $J = 170.8$), 72.9 (d, $J = 26.2$), 59.0 (d, $J = 1.8$), 53.5, 53.3, 47.0 (d, $J = 18.8$), 23.6 (d, $J = 11.5$), 21.7 (d, $J = 24.3$), 17.8, 16.8 (d, $J = 6.9$), 14.1, 12.7, 11.4. ^{19}F NMR (376 MHz, CDCl_3): δ –139.7. IR (cm^{-1}): 3451, 3027, 2965, 2881, 2825, 1495, 1452, 1388, 1371, 1308, 1242, 1168, 1122, 1091, 1064, 1044, 1028, 1013, 930, 898, 867, 841, 804. LC-MS: Exact mass calculated for $[\text{C}_{19}\text{H}_{30}\text{FNO} + \text{H}]^+$: 308.2390 m/z ; found: 308.2361 m/z .

Product 6d. Employing the General Procedure using epoxide 4c (42.0 mg), 6d was obtained as an orange oil (33.8 mg, 75%, $rr > 20:1$). ^1H NMR (500 MHz, CDCl_3): δ 3.47–3.41 (m, 1 H), 3.18 (br s, 1 H), 2.75–2.66 (m, 1 H), 2.58–2.52 (m, 1 H), 2.20–2.09 (m, 1 H), 1.86–1.56 (m, 7 H), 1.55–1.29 (m, 10H), 1.29–1.06 (m, 6 H), 1.04 (s, 3 H), 1.02 (d, $J = 6.9$, 3 H), 0.91 (dd, $J = 7.3$, 7.2, 3 H), 0.84 (dd, $J = 7.4$, 7.5, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 100.7 (d, $J = 171.1$), 72.4 (d, $J = 26.8$), 56.9, 56.5 (d, $J = 2.1$), 52.0, 48.0 (d, $J = 18.2$), 34.2, 33.3, 27.1, 26.6, 26.2, 24.3 (d, $J = 7.8$), 23.8 (d, $J = 11.2$), 21.9 (d, $J = 24.7$), 17.2, 14.5, 11.9, 11.5. ^{19}F NMR (376 MHz, CDCl_3): δ –139.3. IR (cm^{-1}): 2928, 2854, 1451, 1383, 1132, 1091, 1058, 995, 923, 900, 892, 871, 803. LC-MS: Exact mass calculated for $[\text{C}_{18}\text{H}_{34}\text{FNO} + \text{H}]^+$: 300.2703 m/z ; found: 300.2705 m/z .

Product 7c. Employing the General Procedure using epoxide 5c (42.5 mg), 7c was obtained as an orange oil (36.6 mg, 80%, $rr = 94:6$). ^1H NMR (500 MHz, CDCl_3): δ 7.34–7.28 (m, 4 H), 7.28–7.22 (m, 1 H, *para*-H of Bn), 4.17 (d, $J = 13.2$, 1 H), 3.62 (br s, 1 H), 3.04 (d, $J = 13.2$, 1 H), 2.70 (dd, $J = 7.8$, 3.8, 1 H), 2.59–2.53 (m, 1 H), 2.26–2.19 (m, 1 H), 2.01–1.62 (m, 5 H), 1.52–1.44 (m, 1 H), 1.35 (d, $J = 21.8$, 3 H), 1.11 (dd, $J = 7.6$, 7.5, 3 H), 0.99 (ddd, $J = 7.8$, 7.8, 3.3, 3 H). $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3): δ 139.8, 128.5, 128.4, 127.0, 97.2 (d, $J = 175.2$), 74.5 (d, $J = 22.4$), 62.9, 57.1, 47.6, 34.2 (d, $J = 21.4$), 26.1 (d, $J = 1.2$), 21.5 (d, $J = 23.5$), 20.1, 11.4, 8.6 (d, $J = 7.6$). ^{19}F NMR (376 MHz, CDCl_3): δ –156.9. IR (cm^{-1}): 3451, 2965, 2881, 2825, 1495, 1452, 1388, 1371, 1308, 1242, 1168, 1122, 1091, 1064,

1044, 1028, 1013, 951, 930, 898, 867, 841, 804. LC-MS: Exact mass calculated for $[C_{17}H_{26}FNO + H]^+$: 280.2077 m/z ; found: 280.2083 m/z .

■ ASSOCIATED CONTENT

■ Supporting Information

1H , ^{13}C , and ^{19}F NMR spectra of new compounds, and X-ray structures of **4a**, **4g**, **5a**, **5c**, **6a**, **6b**. The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.5b00816.

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Notes

The authors declare no competing financial interest.

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- (15) Synthetic details, characterization, and molecular structures obtained by X-ray structural analysis of compounds are described in the Supporting Information under CCDC nos.: 1000666 (**4a**), 1000667 (**4g**), 1000668 (**5a**), 1000669 (**5c**), 1000670 (**6a**), and 1000671 (**6b**). These data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.
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